

Available online at www.sciencedirect.com

Tetrahedron

Tetrahedron 64 (2008) 3541-3547

www.elsevier.com/locate/tet

From cyclopentadiene to isoxazoline-carbocyclic nucleosides: a rapid access to biological molecules through aza-Diels-Alder reactions

Paolo Quadrelli^{a,*}, Andrea Piccanello^a, Mariella Mella^a, Antonino Corsaro^b, Venerando Pistarà ^b

a Dipartimento di Chimica Organica, Università degli Studi di Pavia, Viale Taramelli, 10 I-27100-Pavia, Italy
^b Dipartimento di Scienze Chimiche, Università degli Studi di Catania, Viale A. Doria, 6 I-95125-Catania, Ital

Received 19 October 2007; received in revised form 10 January 2008; accepted 31 January 2008 Available online 3 February 2008

Abstract

A rapid access to carbocyclic nucleosides containing a fused isoxazoline ring is proposed through the Grieco cycloaddition of cyclopentadiene to iminium salts. The prolific elaboration of the isoxazoline cycloadducts allowed preparation of the target aminols through the unmasking of the hydroxymethylene group at the C3 level of the azanorbornene structure. The heterocyclic aminols are readily converted into nucleosides via the linear construction of purine heterocycles.

© 2008 Elsevier Ltd. All rights reserved.

1. Introduction

Nucleosides are important synthetic targets and the search for analogues containing significantly altered carbohydrate moieties is actively pursued because of their potential antiviral and/or antitumor activities.^{[1](#page-6-0)} A variety of modifications of the nucleoside structures have been made and resulted in different derivatives including di-deoxynucleosides where both hydroxy groups on the sugar ring are removed, 2 carbocyclic and heterocyclic nucleosides where the sugar moiety is replaced with a carbocyclic or a heterocyclic ring, 3 acyclic-nucleosides as well as iso-nucleosides, where the nucleobases are transposed from the 1'-position to the 2'-position of a furanose ring.^{$\bar{4}$ $\bar{4}$ $\bar{4}$} New efforts are constantly made to propose attractive synthetic strategies toward new compounds, with potentially increased biological activities and decreased toxicities.^{[3](#page-6-0)}

Among the various modifications, carbocyclic nucleosides are extensively investigated because the replacement of the furanose ring oxygen by a methylene group results in the stabilized glycosyl bond under acidic conditions and higher

0040-4020/\$ - see front matter © 2008 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2008.01.124

resistance to metabolic enzymes.^{[3,5](#page-6-0)} Ring size modifications as well as the mono and polycyclic nature of the spacer between the nucleobases and the hydroxy group affect the conformational bias as well as the activity and function of the nucleoside analogues, as detailed in various structural studies.^{[6](#page-6-0)} A variety of synthetic approaches are directed to increase the molecular diversity in nucleosidic compounds, which nevertheless must have the features schematically depicted in $1.^7$ $1.^7$

The carbocyclic moiety is normally constituted by a cyclopentane ring but four-, six-, and seven-membered ring are often proposed as alternatives.⁸ The carbocyclic ring is the center of the subsequent structural modifications and can be either saturated or unsaturated and carrying substituents or fused to carbocyclic or heterocyclic rings. A side arm carrying a hydroxy group is always present. This last feature discriminates between the two families of the classical nucleosides $(x=1)$ bearing a hydroxymethylene moiety and the *nor*-nucleosides $(x=0)$ having the hydroxy group directly linked to the

^{*} Corresponding author. Tel.: $+39$ 0382 987315; fax: $+39$ 0382 987323. E-mail address: paolo.quadrelli@unipv.it (P. Quadrelli).

carbocyclic ring. 9 The nucleobases can be of the purine or pyrimidine type and a variety of functionalized structures have been reported, suitable for various functionalizations.

In this context, we have prepared a new class of *nor*nucleosides containing a fused isoxazoline ring by the linear construction of the desired purine and pyrimidine bases on the aminols obtained from the nitrosocarbonyl adducts with cyclopentadiene.[10](#page-6-0) Their activity as potential antiviral agents has been tested against Herpes Simplex virus types 1 and $2¹¹$ $2¹¹$ $2¹¹$ In order to evaluate the effect of the elongation of the hydroxy side chain we have recently proposed a novel approach to useful aminols for the synthesis of carbocyclic nucleosides starting from a convenient source, the 2-azanorborn-5-enes 3. These are readily available through the Grieco cycloaddition of cyclopentadiene 2 with iminium salts^{[12](#page-6-0)} generated in situ under Mannich-like conditions, in a mild and convenient aqueous aza-Diels-Alder (ADA) reaction.^{[13](#page-6-0)} They are quite reactive dipolarophiles toward nitrile oxides 14 and afford exclusively the exo adducts 4. This exclusive exo selectivity of the norbornene-like dipolarophiles has attracted a great deal of attention and is usually attributed to relief of strain, geometric deformation of the double bond (pyramidalization due to π/σ repulsion), and also favorable staggering effects for the exo attack.^{[10d](#page-6-0)} The prolific elaboration of the *exo* isoxazoline cycloadducts 4 allowed the preparation of the target aminols 7 through the intermediates 5 and 6 by using NBS oxidation and subsequent solvolysis for the unmasking of the hydroxymethylene group at the C3 level of the azanorbornene structure (Scheme 1).

On pursuing our studies on nucleoside syntheses, we detail here the first synthesis of a class of racemic purine-carbocyclic nucleosides containing a fused isoxazole ring and having a hydroxymethylene $(HO-CH₂)$ group in the side chain of the carbocyclic unit.

2. Results and discussion

The ADA cycloadduct 3 between cyclopentadiene 2 and the benzyl-iminium salt $[PhCH_2-NH^+=CH_2]$ was transformed into the stereodefined regioisomeric anti aminols of type 7 (Scheme 1). These syntheses have been performed according to the previously described protocols, 14 which nevertheless required a proper tuning for the scale-up necessary for the synthesis of nucleosides in the due amount for the biological tests.

By adapting the known procedures, $5b,15$ the regioisomeric aminols 7a,b have been converted into the pyrimidine derivatives 8a,b through condensation with the 5-amino-4,6-dichloropyrimidine and then into the chloropurines 9a,b with orthoformates under HCl catalysis (Scheme 2). The pyrimidine derivatives 8a,b were obtained in good yields (8a, 72%; 8b 75%) by heating a solution of the aminols 7a,b and 5-amino-4,6-dichloropyrimidine (2 equiv) in n -BuOH at reflux (bp 117 °C) in the presence of an excess of i -Pr₂NEt (5 equiv) for 48 h. The epimeric aminol $7a'$ was also converted into the pyrimidine derivatives $8a'$ (79%) and this latter into the chloropurines $9a'$ under the same conditions.

Scheme 2. (a) 5-Amino-4,6-dichloropyrimidine (2 equiv), i -Pr₂NEt (5 equiv), $n-\text{BuOH}$, Δ , 48 h; (b) HC(OEt)₃/HCl, rt, 8 days.

The structures of $8a,a',b$ rely upon the analytical and spectroscopic data. While the IR spectra of pyrimidines 8a,a',b exhibit the expected bands between $3200-3500$ cm⁻¹ due to the presence of OH, NH, and NH_2 groups, the ${}^{1}H$ NMR spectra were unambiguously consistent for the assigned structures. The spectrum of $8a$ in CD_3COCD_3 showed the pyrimidine ring proton as a singlet at δ 8.03, the NH₂ protons as a broad singlet at δ 4.43 and the OH proton as a triplet at δ 4.54 because of the coupling with the methylene at δ 3.89 (AB syst.), while the NH proton is a doublet at δ 7.01 (*J*= 6.9 Hz). The 5- and 4-isoxazolinic protons are found at δ 5.22 (dd, $J=9.6$, 3 Hz) and 4.29 (dd, $J=9.6$, 3 Hz), respectively, while the cyclopentane protons are at δ 4.85 (m, CH-N), 2.49 (CH-CH₂-O), and 2.29, 1.77 (m, CH₂).

The spectrum of the regioisomeric **8b** is essentially similar, showing the pyrimidine singlet at δ 7.90, the NH₂ protons as a broad singlet at δ 4.39 and OH proton as a broad singlet at δ 4.77 while the corresponding methylene is at δ 3.87 (AB syst.), the NH at δ 6.85 (d, J=6.4 Hz) and the 5- and 4-isoxazolinic protons at δ 5.11 (d, J=9 Hz) and 4.24 (dd, J=9, 3 Hz). The cyclopentane protons are found at δ 4.77 (m, $CH-NH$), 2.46 (m, CH), and 2.35, 1.77 (m, CH₂).

Analogously, the spectrum of the stereoisomeric $8a'$ shows the pyrimidine singlet at δ 7.93, the NH₂ protons as a broad singlet at δ 4.72, and the NH at δ 6.30 (d, J=5 Hz). The OH

proton appears as a triplet at δ 4.65 (J=6 Hz) while the corresponding methylene is at δ 3.71 (AB syst.). The 5- and 4-isoxazolinic protons are shown at δ 5.27 (dd, J=9, 5 Hz) and 4.43 (d, $J=9$ Hz) and the cyclopentane protons are found at δ 3.91 $(m, CH-NH), 2.88$ $(m, CH),$ and 2.02, 1.56 $(m, CH₂)$.

The conversion of the stereoisomeric pyrimidine $8a$, a' , b into the corresponding chloropurines $9a$, a' , b was performed using excess triethyl orthoformate in the presence of catalytic 37% HCl by keeping the reactions at rt for 8 days, in accor-dance with the most frequently reported procedures.^{[16](#page-6-0)} The chloropurines $9a$, a' , b could finally be isolated in good yields $(9a, 87\%, 9a', 82\%$ and $9b, 84\%)$. Contrary to expectations, the reactions performed with the method used for the preparation of the *nor*-nucleosides, $\frac{1}{a}$ in the presence of excess triethyl orthoformate and catalytic p-TsOH, did not take place and the pyrimidine derivatives $8a$,a',b were recovered unchanged.

The chloropurine 9a has been fully characterized spectroscopically. Infrared spectra showed a single broad band at 3320 cm⁻¹ corresponding to the OH absorptions. In the ${}^{1}H$ NMR spectra (CDCl₃) the two N=*CH* protons of the purine rings occur as singlets at δ 8.76 and 8.02 while the 5- and 4-isoxazolinic protons appear as double doublets at δ 5.47 and 4.82 (J_{H4H5} =10 Hz). Similarly, the regioisomeric chloropurine **9b** shows a single broad band at 3373 cm^{-1} corresponding to the OH absorptions in the infrared spectrum and in the 1 H NMR spectra the two N=CH protons of the purine rings occur as singlets at δ 8.80 and 8.27. The 5- and 4-isoxazolinic protons appear as double doublets at δ 5.74 and 4.47 $(J_{\text{H4H5}} = 10 \text{ Hz}).$

The stereoisomeric chloropurine $9a'$ shows a single broad band at 3353 cm⁻¹ corresponding to the OH absorptions in the infrared spectrum and in the H NMR spectra the two $N=CH$ protons of the purine rings occur as singlets at δ 8.80 and 8.13. The 5- and 4-isoxazolinic protons appear at δ 5.67 (dd, J=9, 5 Hz) and 4.93 (d, J=9 Hz).

From the chloro-substituted nucleosides $9a$, a' , b a variety of derivatives can be obtained by nucleophilic substitution.^{[17](#page-6-0)} On heating MeOH solutions of $9a,a',b$ at 50 °C in the presence of an excess of $NH₃$ or other differently substituted amines, the amino derivatives $10a,a',b(A-C)$ could easily be obtained (Scheme 3). Ethoxy derivatives $10a$, a' , b D were directly obtained from pyrimidine derivatives $9a,a',b$ when orthoformate/ H^+ treatment is prolonged for 14 days.

Table 1 reports the chemical yields and physical constants of nucleosides $10a,a',b(A-D)$, which have been fully characterized through their analytical and spectroscopic data. The IR spectra of the adenine derivatives 10aA, 10a'A, and 10bA showed neat and distinctive OH bands (3551, 3399, and 3359 cm⁻¹, respectively) and NH₂ bands (3311, 3143; 3426, 3329, and 3314, 3114 cm^{-1} , respectively). The ¹H NMR spectra showed the characteristic signals of adenine $CH =$ singlets at δ 8.16, 8.06; 8.28, 8.15 and 8.22, 8.19 respectively). The isoxazolinic protons occur as double doublets, because of an additional coupling with the adjacent cyclopentane methines, thus indicating the same conformation in the cyclopentane ring of the chloro-nucleosides $9a, a', b$. The major coupling constant $(J=10 \text{ Hz}$ in **10a,bA** and $J=8.5 \text{ Hz}$ in **10a'A**) is

Scheme 3. (a) RNH₂/MeOH, 50 °C, 24 h; R: see Table 1.

Table 1 Yields and physical constants of purine derivatives

	R	Mp ($°C$) (Solvent)	Yield $(\%)$
9а	C1	>200 dec (Diisopropyl ether)	87
10aA	NH ₂	224 dec (Diisopropyl ether)	89
10aB	NHMe	186–188 (Diisopropyl ether)	92
10aC	NHcPr	Oil	90
10aD	OEt	$130-132$ (<i>n</i> -Hexane/benzene)	84
9a'	C1	>200 dec (<i>n</i> -Hexane/benzene)	82
$10a'$ A	NH ₂	>200 dec (Diisopropyl ether)	88
10a'B	NHMe	$191-193$ (Diisopropyl ether)	95
$10a^{\prime}C$	NHcPr	$170-175$ dec (Diisopropyl ether)	80
10a'D	OEt	$171 - 173$ (Benzene)	77
9b	C1	>200 dec (Diisopropyl ether)	84
10 _b A	NH ₂	>220 dec (<i>n</i> -Hexane/benzene)	72
10 _b B	NHMe	>200 dec (<i>n</i> -Hexane/benzene)	87
10 _b C	NHcPr	>220 dec (<i>n</i> -Hexane/benzene)	80
10 _b D	OEt	180-182 (Benzene)	80

between the H4- and H5-isoxazolinic protons while those between the isoxazolinic protons and the adjacent methines differ in the two regioisomers (10aA, $J=5$ Hz, 10a $'A$, $J=5$ Hz, and 10bA, $J=6$ Hz).

In the amine derivatives $10a,a',b(B,C)$ the IR spectra showed the NH bands in the range $3257-3394$ cm⁻¹. The ¹H NMP spectra showed the characteristic NH signals as ¹H NMR spectra showed the characteristic NH signals as broad singlets at δ 6.87, 6.91 for **10aB,C**, δ 6.83, 6.90 for 10a'B,C, and δ 6.82, 6.92 for 10bB,C. In ethoxy derivatives 10a,a',bD the ¹H NMR spectra showed the OCH₂CH₃ signals, the methylenes as quartets at δ 4.72, 4.71, 4.71 and the methyls as triplets at δ 1.57, 1.55, 1.55, respectively.

These compounds, as well as the pyrimidine derivatives whose synthesis is actually in progress, have been taken as candidates for the biological evaluation against the same viruses in order to have a comparison at the SAR level.

3. Conclusions

The first synthesis of isoxazoline-carbocyclic nucleosides having a hydroxymethylene side chain and a variety of analogues was attained starting from the stereodefined heterocyclic aminols 7a,a',b, which are readily available through exo selective 1,3-dipolar cycloadditions of benzonitrile oxide to 2-azanorborn-5-enes 3 ($R = CH_2-Ph$) and elaboration of the cycloadducts. The stereodefined heterocyclic aminols $7a,a',b$ afford the carbocyclic skeleton for the linear construction of the purine rings. Functionalization of the chloropurines 9a,a',b with a variety of amines extended the synthetic potential of this strategy allowing for a fine tuning of their biological and antiviral activity as well as comparison with the corresponding nor-nucleosides.

Biological evaluation of the newly obtained compounds 10a,a',b is actually in progress.

4. Experimental

4.1. General

All melting points are uncorrected. Elemental analyses were done on a C. Erba 1106 elemental analyzer. IR spectra (Nujol mulls) were recorded on an FT-IR Perkin-Elmer RX-1. 1 H and 13 C NMR spectra were recorded on a Bruker AVANCE 300 in the specified deuterated solvents. Chemical shifts are expressed in parts per million from internal tetramethylsilane (δ) . UV-vis spectra were recorded on a UV Perkin-Elmer LAMBDA 16 spectrophotometer using acetonitrile as solvent. HPLC analyses were carried out by means of a WATERS 1525 instrument equipped with an UV 2487 detector (λ =266 nm) both controlled by Breeze[™] software and a RP C-18 Intersil ODS-2 column; a mixture of H_2O/CH_3CN 50:50 was used as eluant. Column chromatography and TLC: silica gel $60 (0.063-0.200 \text{ mm})$ (Merck); eluant chloroform. The identification of samples from different experiments was secured by mixed mps and superimposable IR spectra.

4.2. Materials

ADA cycloadduct 3 has been prepared according to Grieco's procedure.^{[12](#page-6-0)} Benzhydroximoyl chloride was prepared according to the reported method.^{[18](#page-6-0)} The cycloaddition of BNO to ADA cycloaddutct 3 was performed according to the procedure described in the previous study, as well as the oxidation of the regioisomeric 1,3-dipolar cycloadducts with m-CPBA and the Polonovski rearrangement of the N-oxides with acetic anhydride.^{[14](#page-6-0)}

The N-acetyl derivatives 4 were then dissolved in $CCl₄$ (40 mL/g) and 1.5 equiv NBS was added along with 15 mol % of AIBN. The mixtures were refluxed at reflux monitoring the reaction by TLC until the starting materials have disappeared $(2-3 h)$. The solutions were cooled down to ambient and then to lower temperature in an ice-bath to ensure the maximum separation of succinimide. From the filtrate organic phases the solvent was removed upon evaporation and the crude residues were submitted to chromatographic separation on silica gel by eluting initially with $CHCl₃$ and $CHCl₃$ / MeOH 9:1 afterward. The bromo-cycloadducts 5 were collected along with the products of their solvolysis, the acetals, and the corresponding aldehydes. The 3-bromine derivatives 5 were treated with excess $NaHCO₃$ in refluxing EtOH for several days to promote their solvolysis to the aldehydes of type 6. Reduction of these latters and deprotection of the amino group afforded the desired aminols $7.^{14}$ $7.^{14}$ $7.^{14}$

4.3. Synthesis of the pyrimidine derivatives $8a,a',b$

To aminols $7a,a',b$ (1.70 g, 7.32 mmol) dissolved in n-BuOH (75 mL), 5-amino-4,6-dichloropyrimidine (2.55 g, 15.5 mmol) and i -Pr₂NEt (4.02 g, 31.1 mmol) were added. The mixtures were heated at 117 °C with stirring for 48 h. The cooled solutions were evaporated to dryness, taken up in $CH₂Cl₂$, washed with brine, and dried over anhydrous Na2SO4. The crude residues were then submitted to column chromatography to separate the excess of amino-pyrimidine from adducts $8a-c$, which were isolated in 72, 79, and 75% yields, respectively.

4.3.1. Compound 8a

Yield: 1.90 g, 72% as white crystals from ethanol, mp $175-$ 180 °C; [Found C, 56.6; H, 5.0; N, 19.3. C₁₇H₁₈N₅O₂Cl (MW=359.82) requires C, 56.75; H, 5.04; N, 19.46%]; v_{max} (Nujol) 3411, 3348, 3303, 3257, 1658 cm⁻¹; δ_H (300 MHz, CD_3COCD_3) 8.03 (1H, s, CH=N), 7.91 (2H, m, Ph), 7.39 (3H, m, Ph), 7.01 (1H, d, J 6.9 Hz, NH), 5.22 (1H, dd, J 9.6, 3 Hz, H5-isoxaz.), 4.85 (1H, m, CH-NH), 4.54 (1H, t, J 4.5 Hz, CH₂ $-OH$), 4.43 (2H, br s, NH₂), 4.29 (1H, dd, J 9.6, 3 Hz, H4-isoxaz.), 3.89 (2H, m, CH_2 –OH), 1.59 (1H, m, CH), 2.29 (1H, m, H-CH), 1.77 (1H, dt, J 13, 5 Hz, H-CH); δ_C (75 MHz, CD₃COCD₃) 158.6, 153.7, 148.3, 140.9, 130.9, 130.7, 129.8, 128.5, 124.7, 90.6, 64.3, 61.0, 57.7, 50.7, 36.3.

$4.3.2.$ Compound $8a'$

Yield: 1.98 g, 75% as white crystals from ethanol, mp>210 °C (dec); [Found C, 56.7; H, 5.0; N, 19.5. $C_{17}H_{18}N_5O_2Cl$ (MW=359.82) requires C, 56.75; H, 5.04; N, 19.46%]; v_{max} (Nujol) 3412, 3368, 3300, 3157, 1646 cm⁻¹; δ_{H} $(300 \text{ MHz}, \text{CD}_3\text{COCD}_3)$ 8.09 (2H, m, Ph), 7.93 (1H, s, CH= N), 7.46 (3H, m, Ph), 6.30 (1H, d, J 5 Hz, NH), 5.27 (1H, dd, J 9, 5 Hz, H5-isoxaz.), 4.72 (2H, br s, NH₂), 4.65 (1H, t, J 6 Hz, CH_2 -OH), 4.43 (1H, d, J 9 Hz, H4-isoxaz.), 3.91 (1H, m, CH-NH), 3.71 (2H, AB syst., J 13 Hz, CH₂-OH), 2.88 (1H, m, CH), 2.02 (1H, m, H-CH), 1.56 (1H, m, H-CH); δ_C $(75 \text{ MHz}, \text{ CD}_3\text{COCD}_3)$ 157.9, 153.1, 147.7, 139.9, 130.9, 130.7, 129.9, 128.6, 125.0, 87.8, 62.1, 60.7, 57.6, 49.2, 34.3.

4.3.3. Compound 8b

Yield: 2.08 g, 79% as white crystals from ethanol, mp $118-$ 120 °C; [Found C, 56.7; H, 5.1; N, 19.5. C₁₇H₁₈N₅O₂Cl (MW=359.82) requires C, 56.75; H, 5.04; N, 19.46%]; v_{max} (Nujol) 3391, 3327, 3283, 3233, 1654 cm⁻¹; δ_H (300 MHz, CD_3COCD_3) 7.90 (1H, s, CH=N), 7.83 (2H, m, Ph), 7.46 (3H, m, Ph), 6.85 (1H, d, J 6.4 Hz, NH), 5.11 (1H, d, J 9 Hz, H5-isoxaz.), 4.77 (1H+1H, m, CH-NH+OH), 4.39 (2H, br s, NH2), 4.24 (1H, dd, J 9, 3 Hz, H4-isoxaz.), 3.87 (2H, m, CH_2 -OH), 2.46 (1H, m, CH), 2.35 (1H, m, H-CH),

1.77 (1H, m, H-CH); δ_C (75 MHz, CD₃COCD₃) 154.3, 147.1, 141.3, 130.9, 124.3, 123.6, 123.2, 121.6, 118.0, 87.6, 58.4, 53.2, 47.9, 41.0, 27.9.

4.4. Construction of the purine nucleosides $9a,a',b$

To a solution of pyrimidine derivatives θa , a ['], b (0.532 g, 1.48 mmol) in triethyl orthoformate (25 mL), a catalytic amount of HCl was added. The reaction was stirred at rt for 8 days. After this period of time, the excess orthoformate was hydrolyzed and the water phase extracted with CH_2Cl_2 and dried over anhydrous $Na₂SO₄$. After evaporation of the solvent, the residue was taken up with diisopropyl ether and the chloropurine derivatives $9a$,a',b crystallized.

4.4.1. Compound 9a

Yield: 0.48 g, 87% as white crystals from diisopropyl ether, mp > 200 °C (dec); [Found C, 58.4; H, 4.4; N, 19.0. $C_{18}H_{16}N_5O_2Cl$ (MW=369.82) requires C, 58.46; H, 4.36; N, 18.94%]; v_{max} (Nujol) 3320, 1654, 1560 cm⁻¹; δ_{H} (300 MHz, CDCl₃) 8.76 (1H, s, CH=N), 8.02 (1H, s, CH=N), 7.29 (5H, m, Ph), 5.47 (1H, dd, J 10, 6 Hz, H5-isoxaz.), 4.97 (1H, m, CH-N), 4.82 (1H, dd, J 10, 7 Hz, H4-isoxaz.), 4.01 (2H, m, CH₂-OH), 2.58 (2H, m, CH₂), 2.43 (1H, m, CH); δ_C (75 MHz, CDCl3) 159.3, 156.6, 151.8, 144.5, 144.3, 129.0, 128.5, 127.6, 126.7, 122.3, 88.0, 62.9, 61.0, 57.9, 48.0, 34.3.

4.4.2. Compound $9a'$

Yield: 0.45 g, 82% as white crystals from benzene/*n*-hexane, mp > 200 °C (dec); [Found C, 58.5; H, 4.3; N, 18.9. $C_{18}H_{16}N_5O_2Cl$ (MW=369.82) requires C, 58.46; H, 4.36; N, 18.94%]; v_{max} (Nujol) 3353, 1655, 1580 cm⁻¹; δ_{H} $(300 \text{ MHz}, \text{ CDCl}_3)$ 8.80 (1H, s, CH=N), 8.13 (1H, s, CH=N), 7.68 (2H, m, Ph), 7.43 (3H, m, Ph), 5.67 (1H, dd, J 9, 5 Hz, H5-isoxaz.), 5.15 (1H, d, J 6 Hz, CH-N), 4.93 (1H, d, J 9 Hz, H4-isoxaz.), 4.00 (2H, m, CH_2 -OH), 3.13 (1H, m, CH), 2.24 (2H, m, CH₂); δ_C (75 MHz, CDCl₃) 156.5, 151.9, 151.3, 144.0, 134.0, 130.7, 129.1, 127.5, 126.9, 88.3, 61.2, 61.0, 58.9, 47.6, 34.0.

4.4.3. Compound 9b

Yield: 0.46 g, 84% as white crystals from diisopropyl ether, mp>200 °C (dec); [Found C, 58.5; H, 4.5; N, 19.1. $C_{18}H_{16}N_5O_2Cl$ (MW=369.82) requires C, 58.46; H, 4.36; N, 18.94%]; v_{max} (Nujol) 3373, 1597, 1561 cm⁻¹; δ_{H} $(300 \text{ MHz}, \text{ CDCl}_3)$ 8.80 (1H, s, CH=N), 8.27 (1H, s, CH=N), 7.78 (2H, m, Ph), 7.48 (3H, m, Ph), 5.74 (1H, dd, J 10, 6 Hz, H5-isoxaz.), 4.99 (1H, m, CH-N), 4.47 (1H, dd, J 10, 5 Hz, H4-isoxaz.), 3.91 (2H, m, CH₂-OH), 2.64 (2H, m, CH₂), 2.47 (1H, m, CH); δ_C (75 MHz, CDCl₃) 159.9, 151.6, 151.4, 144.6, 132.4, 130.5, 128.9, 128.0, 127.3, 89.1, 64.3, 63.9, 53.1, 43.9, 32.9.

4.5. Syntheses of the amino derivatives $10a,a',b$

General method: Solutions of chloro-nucleosides 9a,a',b (30 mg, 0.08 mmol) in MeOH (2 mL) were saturated with ammonia or other gaseous amines and kept in a sealed tube at 50° C for 24 h. In the case of liquid amines, an excess (50 equiv) was added to the solutions. The solutions are then cooled and concentration of the solutions afforded oily residues from which the amino derivatives were crystallized from the proper solvent. [Table 1](#page-2-0) reports the physical constants (solvent of crystallization) and yields (determined by HPLC analyses) of the amino nucleosides $10a.a', b(A-C)$. Ethoxy derivatives 10a,a',bD were obtained upon extended reaction with ethyl orthoformate up to 14 days and further addition of drops of 37% HCl and usual work-up.

4.5.1. Compound 10aA

Yield: 25 mg, 89% as white crystals from diisopropyl ether, mp 224 °C (dec); [Found C, 61.3; H, 5.1; N, 23.7. C₁₈H₁₈N₆O₂ (MW=350.37) requires C, 61.70; H, 5.18; N, 23.99%]; ν_{max} (Nujol) 3551, 3311, 3143, 1666 cm⁻¹; δ_{H} (300 MHz, CD_3COCD_3) 8.16 (1H, s, CH=N), 8.06 (1H, s, CH=N), 7.25 (5H, m, Ph), 6.63 (2H, br s, NH2), 5.36 (1H, dd, J 10, 5 Hz, H5-isoxaz.), 5.02 (2H, m, CH-N and H4-isoxaz.), 4.07 (1H, br s, OH), 3.84 (2H, m, CH_2 –OH), 2.42 (3H, m, CH₂ and CH); δ_C (75 MHz, CD₃COCD₃) 165.2, 159.2, 153.6, 148.6, 141.4, 138.4, 131.0, 129.9, 129.8, 128.0, 89.4, 63.4, 61.2, 58.3, 50.0, 36.4.

4.5.2. Compound 10aB

Yield: 27 mg, 92% as white crystals from diisopropyl ether, mp 186–188 °C; [Found C, 62.3; H, 5.5; N, 23.0. C₁₉H₂₀N₆O₂ (MW=364.40) requires C, 62.62; H, 5.53; N, 23.06%]; v_{max} (Nujol) 3402, 3257, 1625 cm⁻¹; δ_H (300 MHz, CD₃COCD₃) 8.23 (1H, s, CH=N), 8.01 (1H, s, CH=N), 7.31 (5H, m, Ph), 6.87 (1H, br s, NH), 5.36 (1H, dd, J 10, 5 Hz, H5-isoxaz.), 5.03 (2H, m, CH-N and H4-isoxaz.), 4.11 (1H, br s, OH), 3.83 (2H, m, CH_2 –OH), 2.40 (3H, m, CH₂ and CH), 2.06 (3H, d, J 7 Hz, CH₃); δ_C (75 MHz, CD₃COCD₃) 159.0, 153.8, 140.8, 131.0, 130.2, 129.8, 128.5, 128.0, 89.5, 63.3, 61.2, 58.3, 49.9, 36.5, 27.7.

4.5.3. Compound $10aC$

Yield: 28 mg, 90% as yellowish oil; [Found C, 64.5; H, 5.6; N, 21.5. $C_{21}H_{22}N_6O_2$ (MW=390.43) requires C, 64.60; H, 5.68; N, 21.53%]; ν_{max} (Nujol) 3297, 1622, 1572 cm⁻¹; δ_{H} $(300 \text{ MHz}, \text{CD}_3\text{COCD}_3)$ 8.24 (1H, s, CH=N), 8.02 (1H, s, CH=N), 7.23 (5H, m, Ph), 6.91 (1H, br s, NH), 5.32 (1H, dd, J 10, 5 Hz, H5-isoxaz.), 5.03 (2H, m, CH-N and H4-isoxaz.), 3.83 (2H, m, CH-NH and CH), 3.73 (2H, m, O-CH₂), 2.70 (2H, m, CH₂), 0.89 (2H, m, CH₂), 0.63 (2H, m, CH₂); δ_C (75 MHz, CD₃COCD₃) 162.0, 152.3, 139.6, 129.6 129.5, 128.4, 126.6, 122.3, 88.1, 62.0, 59.8, 57.0, 48.6, 35.0, 22.0, 6.2, 5.3.

4.5.4. Compound 10aD

Yield: 25 mg, 84% as white crystals from *n*-hexane/ benzene, mp 130-132 °C; [Found C, 63.2; H, 5.6; N, 18.5. $C_{20}H_{21}N_5O_3$ (MW=379.41) requires C, 63.31; H, 5.58; N, 18.46%]; v_{max} (Nujol) 3292, 1598, 1575 cm⁻¹; δ_{H} $(300 \text{ MHz}, \text{ CDCl}_3)$ 8.52 (1H, s, CH=N), 7.75 (1H, s,

CH=N), 7.36 (2H, m, Ph), 7.25 (3H, m, Ph), 5.46 (1H, dd, J 10, 5 Hz, H5-isoxaz.), 4.88 (1H, m, CH-N), 4.81 (2H, dd, J 10, 3 Hz, H4-isoxaz.), 4.72 (2H, q, J 7 Hz, O-CH₂), 4.00 (2H, br s, CH_2 –OH), 2.63 (2H, m, CH₂), 2.48 (1H, m, CH), 1.57 (3H, t, J 7 Hz, CH₃); δ_C (75 MHz, CDCl₃) 161.1, 158.2, 151.9, 151.1, 141.6, 130.4 128.9, 128.2, 127.8, 126.8, 88.9, 63.4, 63.3, 61.3, 57.8, 47.9, 34.6, 14.4.

$4.5.5.$ Compound $10a'A$

Yield: 25 mg, 88% as white crystals from diisopropyl ether, mp>200 °C (dec); [Found C, 61.5; H, 5.2; N, 23.8. $C_{18}H_{18}N_6O_2$ (MW=350.37) requires C, 61.70; H, 5.18; N, 23.99%]; v_{max} (Nujol) 3426, 3399, 3329, 1654 cm⁻¹; δ_{H} $(300 \text{ MHz}, \text{ DMSO})$ 8.28 (1H, s, CH=N), 8.15 (1H, s, CH=N), 7.70 (2H, m, Ph), 7.42 (3H, m, Ph), 5.45 (1H, dd, J 8.5, 5 Hz, H5-isoxaz.), 4.95 (2H, m, CH-N and H4-isoxaz.), 4.68 (2H, br, NH₂), 3.70 and 3.51 (2H, AB syst., CH_2 –OH), 2.81 (1H, m, CH), 1.98 and 1.72 (2H, m, CH₂); δ_c (75 MHz, DMSO) 156.5, 155.9, 152.2, 149.2, 139.6, 130.1, 129.0, 128.2, 126.9, 119.1, 87.2, 59.8, 58.5, 57.8, 47.7, 34.6.

$4.5.6.$ Compound $10a'B$

Yield: 28 mg, 95% as white crystals from diisopropyl ether, mp 191–193 °C; [Found C, 62.4; H, 5.4; N, 23.1. C₁₉H₂₀N₆O₂ (MW=364.40) requires C, 62.62; H, 5.53; N, 23.06%]; v_{max} (Nujol) 3318, 1630 cm⁻¹; δ_H (300 MHz, CD₃COCD₃) 8.29 $(1H, s, CH=N), 8.11$ (1H, s, CH=N), 7.84 (2H, m, Ph), 7.44 (3H, m, Ph), 6.83 (1H, br, NH), 5.57 (1H, dd, J 9, 5 Hz, H5-isoxaz.), 5.13 (1H, d, J 6 Hz, CH-N), 5.03 (1H, dt, J 9, 1 Hz, H4-isoxaz.), 3.94 and 3.74 (1H+1H, AB syst., CH_2 -OH), 3.16 (1H, br s, OH), 3.05 (1H, m, CH), 2.82 (3H, d, J 8 Hz, N-CH₃), 2.25 and 1.95 (1H+1H, m, CH₂); δ_C (75 MHz, CD₃COCD₃) 157.9, 153.9, 140.2, 131.2, 130.1, 128.3, 89.1, 62.0, 61.9, 60.7, 59.7, 49.6, 35.9.

4.5.7. Compound $10a'C$

Yield: 25 mg, 80% as white crystals from diisopropyl ether, mp 170-175 °C (dec); [Found C, 64.4; H, 5.5; N, 21.4. $C_{21}H_{22}N_6O_2$ (MW=390.43) requires C, 64.60; H, 5.68; N, 21.53%]; v_{max} (Nujol) 3328, 1655, 1618 cm⁻¹; δ_{H} $(300 \text{ MHz}, \text{CD}_3\text{COCD}_3)$ 8.30 (1H, s, CH=N), 8.13 (1H, s, CH=N), 7.84 (2H, m, Ph), 7.43 (3H, m, Ph), 6.90 (1H, br s, NH), 5.57 (1H, dd, J 9, 5 Hz, H5-isoxaz.), 5.13 (1H, d, J 6.6 Hz, CH-N), 5.03 (1H, d, J 9 Hz, H4-isoxaz.), 3.95 (1H, m, $CH-NH$), 3.73 (2H, m, O-CH₂), 3.16 (1H, br s, OH), 3.07 (1H, m, CH), 2.08 and 1.92 (2H, m, CH2), 0.83 (2H, m, CH₂), 0.72 (2H, m, CH₂); δ_C (75 MHz, CD₃COCD₃) 153.8, 152.5, 151.2, 140.4, 134.6, 131.2 130.1, 128.4, 89.1, 62.0, 60.7, 59.7, 49.6, 35.9, 25.0, 7.6, 7.3.

$4.5.8.$ Compound $10a'D$

Yield: 23 mg, 77% as white crystals from benzene, mp 171-173 °C; [Found C, 63.1; H, 5.5; N, 18.4. C₂₀H₂₁N₅O₃ (MW=379.41) requires C, 63.31; H, 5.58; N, 18.46%]; v_{max} (Nujol) 3348, 1596 cm⁻¹; δ_H (300 MHz, CDCl₃) 8.55 (1H, s, CH=N), 7.88 (1H, s, CH=N), 7.72 (2H, m, Ph), 7.44 (3H, m, Ph), 5.66 (1H, dd, J 9, 5 Hz, H5-isoxaz.), 5.11 (1H,

d, J 6 Hz, CH-N), 4.93 (1H, d, J 9 Hz, H4-isoxaz.), 4.71 (2H, q, J 7 Hz, O-CH₂), 4.00 (2H, m, CH₂-OH), 3.15 (1H, m, CH), 2.18 (2H, m, CH₂), 1.55 (3H, t, J 7 Hz, CH₃); δ_C (75 MHz, CDCl₃) 161.0, 156.8, 152.1, 151.6, 140.7, 130.5 129.1, 127.7, 127.0, 122.2, 88.6, 63.2, 61.4, 60.4, 59.0, 47.5, 34.0, 14.4.

4.5.9. Compound 10bA

Yield: $(20 \text{ mg}, 72\%)$ as white crystals from *n*-hexane/benzene, mp > 220 °C (dec); [Found C, 61.5; H, 5.2; N, 23.7. $C_{18}H_{18}N_6O_2$ (MW=350.37) requires C, 61.70; H, 5.18; N, 23.99%]; v_{max} (Nujol) 3359, 3314, 3114, 1673, 1603 cm⁻¹; $\delta_{\rm H}$ (300 MHz, CD₃COCD₃) 8.22 (1H, s, CH=N), 8.19 (1H, s, CH=N), 7.86 (2H, m, Ph), 7.50 (3H, m, Ph), 6.52 (2, br s, NH2), 5.84 (1H, dd, J 10, 6 Hz, H5-isoxaz.), 4.98 (1H, m, CHeN), 4.45 (1H, dd, J 10, 6 Hz, H4-isoxaz.), 4.26 (1H, t, J 5 Hz, OH), 3.79 (2H, m, CH₂-OH), 2.53 (2H, m, CH₂), 2.36 (1H, m, CH); δ_C (75 MHz, CD₃COCD₃) 161.5, 158.9, 157.6, 153.8, 147.6, 141.2, 136.7, 131.2, 130.0, 128.7, 90.3, 64.7, 54.1, 49.3, 34.9, 20.0.

4.5.10. Compound 10bB

Yield: 25 mg, 87% as white crystals from n -hexane/benzene, mp > 200 °C (dec); [Found C, 62.5; H, 5.4; N, 23.2. $C_{19}H_{20}N_6O_2$ (MW=364.40) requires C, 62.62; H, 5.53; N, 23.06%]; v_{max} (Nujol) 3387, 1634, 1576 cm⁻¹; δ_{H} $(300 \text{ MHz}, \text{CD}_3\text{COCD}_3)$ 8.29 (1H, s, CH=N), 8.14 (1H, s, CH=N), 7.87 (2H, m, Ph), 7.49 (3H, m, Ph), 6.82 (1H, br, NH), 5.84 (1H, dd, J 10, 6 Hz, H5-isoxaz.), 4.99 (1H, m, CH-N), 4.46 (1H, dd, J 10, 4 Hz, H4-isoxaz.), 4.30 (1H, br, OH), 3.78 (2H, m, CH₂-OH), 2.81 (3H, d, J 7 Hz, CH₃-NH), 2.51 (2H, m, CH₂), 2.37 (1H, m, CH); δ_C (75 MHz, CD3COCD3) 161.0, 153.8, 140.7, 131.2, 130.5, 130.0, 128.7, 90.4, 64.7, 64.1, 54.1, 45.8, 35.0, 27.5.

4.5.11. Compound 10bC

Yield: 25 mg , 80% as white crystals from *n*-hexane/ benzene, mp>220 °C (dec); [Found C, 64.5; H, 5.6; N, 21.5. $C_{21}H_{22}N_6O_2$ (MW=390.43) requires C, 64.60; H, 5.68; N, 21.53%]; v_{max} (Nujol) 3394, 1619, 1572 cm⁻¹; δ_{H} $(300 \text{ MHz}, \text{CD}_3\text{COCD}_3)$ 8.30 (1H, s, CH=N), 8.16 (1H, s, CH=N), 7.86 (2H, m, Ph), 7.47 (3H, m, Ph), 6.92 (1H, br s, NH), 5.83 (1H, dd, J 10, 6 Hz, H5-isoxaz.), 4.98 (1H, m, CH-N), 4.45 (1H, dd, J 10, 6 Hz, H4-isoxaz.), 4.29 (1H, br, OH), 3.77 (2H, m, CH_2 -OH), 2.51 (2H, m, CH₂), 2.33 (1H, m, CH), 2.10 (1H, m, CH-NH), 0.83 (2H, m, CH₂), 0.73 (2H, m, CH₂); δ_C (75 MHz, CD₃COCD₃) 161.0, 153.7, 140.9, 131.2, 130.5, 130.0, 128.7, 90.3, 64.8, 64.1, 54.1, 45.8, 34.9, 23.6, 7.6, 6.7.

4.5.12. Compound 10bD

Yield: 24 mg, 80% as white crystals from benzene, mp 180-182 °C; [Found C, 63.2; H, 5.6; N, 18.5. C₂₀H₂₁N₅O₃ (MW=379.41) requires C, 63.31; H, 5.58; N, 18.46%]; v_{max} (Nujol) 3294, 1597, 1575 cm⁻¹; δ_H (300 MHz, CDCl₃) 8.55 (1H, s, CH=N), 8.01 (1H, s, CH=N), 7.77 (2H, m, Ph), 7.48 (3H, m, Ph), 5.71 (1H, dd, J 10, 5 Hz, H5-isoxaz.),

4.96 (1H, m, CH-N), 4.71 (2H, q, J 7 Hz, O-CH₂), 4.47 (1H, dd, J 10, 5 Hz, H4-isoxaz.), 3.90 (2H, m, CH₂-OH), 2.94 (1H, br, OH), 2.67 (2H, m, CH₂), 2.49 (1H, m, CH), 1.55 (3H, t, J 7 Hz, CH₃); δ_C (75 MHz, CDCl₃) 160.0, 151.7, 141.6, 130.3, 128.8, 128.2, 127.3, 89.8, 64.6, 63.2, 53.5, 44.2, 33.1, 30.8, 14.4.

Acknowledgements

Financial support by University of Pavia (FAR) and MIUR (PRIN 2005) is gratefully acknowledged.

References and notes

- 1. (a) Hnang, W.; Miller, M. J.; De Clercq, E.; Balzarini, J. Org. Biomol. Chem. 2007 , 5, 1164-1166 and references therein; (b) Mizuno, Y. The Organic Chemistry of Nucleic Acids; Kadansha: Tokyo, 1986; (c) Ueda, T. Chemistry of Nucleosides and Nucleotides; Townsend, Ed.; Plenum: New York, NY, 1988; Vol. 1, Chapter 1; (d) Srivasta, P. C.; Robins, R. K.; Meyer, R. B., Jr. Chemistry of Nucleosides and Nucleotides; Townsend, Ed.; Plenum: New York, NY, 1988; Vol. 1, Chapter 2; (e) Revenkar, G. R.; Robins, R. K. Chemistry of Nucleosides and Nucleotides; Townsend, Ed.; Plenum: New York, NY, 1988; Vol. 2, Chapter 4; (f) Bloomfield, V. A.; Crothers, D. M.; Tinoco, I., Jr. Nucleic Acids; University Science Books: Sausalito, CA, 2000; pp 13-43.
- 2. (a) Mishra, R.; Misra, K. Chem. Lett. $2007, 36, 768-769$; (b) Zhong, M.; Nowak, I.; Robins, M. J. J. Org. Chem. 2006, 71, 7773-7779; (c) Li, N.-S.; Piccirilli, J. A. Synthesis 2005, 2865-2870; (d) Peyrane, F.; Clivio, P. Org. Biomol. Chem. 2005, 3, 1685-1689.
- 3. Jeong, L. S.; Lee, J. A. Antiviral Chem. Chemother. 2004, 15, 235-250 and references therein.
- 4. (a) Jiang, C.; Li, B.; Guan, Z.; Zhenjun, Z.; Liangren, L. Bioorg. Med. Chem. 2007, 15, 3019-3025; (b) Chun, B.-K.; Wang, P.; Hassan, A.; Du, J.; Tharnish, P. M.; Murakami, E.; Stuyer, L.; Otto, M. J.; Schinazi, R. F.; Watanabe, K. A. Nucleosides, Nucleotides Nucleic Acids 2007, 26, 83-97; (c) Yoshimura, Y.; Asami, K.; Matsui, H.; Tanaka, H.; Takahata, H. Org. Lett. 2006, 8, 6015-6018; (d) Alvarez de Cienfuegos, L.; Mota, A. J.; Robles, R. Org. Lett. 2005, 7, 2161-2164; (e) Nair, V. Recent Advances in Nucleosides: Chemistry and Chemotherapy; Chu, C. K., Ed.; Elsevier: Amsterdam, 2002; pp 149-166.
- 5. (a) Rajappan, V. P.; Yin, X.; Schneller, S. W. Tetrahedron 2002, 58, 9889-9895; (b) Ishikura, M.; Murakami, A.; Katagiri, N. Org. Biomol. Chem. 2003, 1, 452-453; (c) Jeong, L. S.; Yoo, S. J.; Lee, K. M.; Koo, M. J.; Choi, W. J.; Kim, H. O.; Park, J. G.; Lee, S. K.; Chun, M. W. J. Med. Chem. 2003, 46, 201-202; (d) Li, F.; Brogan, J. B.; Gage, J. L.; Zhang, D.; Miller, M. J. J. Org. Chem. 2004, 69, 4538-4540; (e) Lee, W.; Miller, M. J. J. Org. Chem. 2004, 69, 4516-4519; (f) Zhou, W.; Gumina, G.; Chong, Y.; Wang, J.; Schinazi, R. F.; Chu, C. K. J. Med. Chem. 2004, 47, 3399-3408; (g) Kim, H. O.; Baek, H. W.; Moon, H. R.; Kim, D.-K.; Chun, M. W.; Jeong, L. S. Org. Biomol. Chem. 2004, 2, 1164-1168; (h) Kifli, N.; Htar, T. T.; De Clerq, E.; Balzarini, J.; Simons, C. Bioorg. Med. Chem. 2004, 12, 3247-3257; (i) Yin, X.-O.; Schneller, S. W. Tetrahedron 2005, 61, 1839–1843; (j) Roy, A.; Serbessa, T.; Schneller, S. W. Bioorg. Med. Chem. 2006, 14, 4980-4986; (k) Ye, W.; Schneller, S. W. J. Org. Chem. 2006, 71, 8641-8643.
- 6. (a) Estevez, L.; Gonzalez Moa, M. J.; Teran, C.; Mosquera, R. A. Tetrahedron 2007 , 63 , $717-726$ and references therein; (b) Perez-Castrp, I.; Caamaño, O.; Garcia, M. D.; Gomez, G.; Fernandez, F.; Ferreira, J.; Lopez, C. Synthesis 2007, 2621-2626.
- 7. (a) Merino, P. Curr. Med. Chem. Anti-Infect. Agents 2002 , 1, 389-411; (b) Crimmins, M. T. Tetrahedron 1998, 54, 9229-9272; (c) Agrofoglio, L.; Suhas, E.; Farese, A.; Condom, R.; Challand, S. R.; Earl, R. A.; Guedj, R. Tetrahedron 1994, 50, 10611-10670.
- 8. (a) Bodnar, B. S.; Miller, M. J. J. Org. Chem. 2007, 72, 3929-3932; (b) Wang, J.; Froeyen, M.; Herdewijn, P. Adv. Antiviral Drug Des. 2004, 4, 119-145; (c) Miller, M. J. Proceedings of the International Symposium on Advances in Synthetic, Combinatorial and Medicinal Chemistry, Moscow, Russia, May 5-8, 2004; T14 and P140. (d) Vogt, P. F.; Miller, M. J. Tetrahedron 1998, 54, 1317-1348; (e) Boger, L.; Weinreb, S. M. Hetero Diels-Alder Methodology in Organic Synthesis; Academic: San Diego, CA, 1987.
- 9. (a) Sato, T.; Aoyagi, S.; Kibayashi, C. Org. Lett. 2003, 5, 3839-3842; (b) Momiyama, N.; Yamamoto, H. Org. Lett. 2002, 4, 3579-3582; (c) Flower, K. R.; Lightfoot, A. P.; Wan, H.; Whiting, A. J. Chem. Soc., Perkin Trans. 1 2002, 2058-2064; (d) Iwasa, S.; Fakhruddin, A.; Tsukamoto, Y.; Kameyama, M.; Nishiyama, H. Tetrahedron Lett. 2002, 43, 6159-6161; (e) Ware, R. W., Jr.; Day, C. S.; King, S. B. J. Org. Chem. 2002, 67, 6174-6180; (f) Chow, C. P.; Shea, K. J.; Sparks, S. M. Org. Lett. 2002, 4, 2637-2640; (g) Surman, M. D.; Mulvihill, M. J.; Miller, M. J. Tetrahedron Lett. 2002, 43, 1131-1134; (h) Surman, M. D.; Mulvihill, M. J.; Miller, M. J. J. Org. Chem. 2002, 67, 4115-4121; (i) Iwasa, S.; Tajima, K.; Tsushima, S.; Nishiyama, H. Tetrahedron Lett. 2001, 42, 5897-5899.
- 10. (a) Quadrelli, P.; Gamba Invernizzi, A.; Caramella, P. Tetrahedron Lett. 1996, 37, 1909-1996; (b) Quadrelli, P.; Mella, M.; Gamba Invernizzi, A.; Caramella, P. Tetrahedron 1999, 55, 10497-10510; (c) Quadrelli, P.; Mella, M.; Carramella, P. Tetrahedron Lett. 1999, 40, 797-800; (d) Quadrelli, P.; Mella, M.; Paganoni, P.; Caramella, P. Eur. J. Org. Chem. 2000, 2613-2622; (e) Quadrelli, P.; Fassardi, V.; Cardarelli, A.; Caramella, P. Eur. J. Org. Chem. 2002, 2058-2065 and references therein.
- 11. (a) Quadrelli, P.; Scrocchi, R.; Caramella, P.; Rescifina, A.; Piperno, A. Tetrahedron 2004, 60, 3643-3651; (b) Quadrelli, P.; Piccanello, A.; Caramella, P.; Piperno, A.; Corsaro, A.; Pistarà, V. Bioorg. Med. Chem., manuscript in preparation.
- 12. Grieco, P. A.; Larsen, S. D. Org. Synth. 1990, 68, 206-209.
- 13. (a) Grieco, P. A.; Bahsas, A. J. Org. Chem. 1987, 52, 5746–5749; (b) Larsen, S. D.; Grieco, P. A. J. Am. Chem. Soc. 1985, 107, 1768-1769; (c) Rodriguez-Borges, J. E.; Garcia-Mera, X.; Fernandez, F.; Lopez, V. H. C.; Magolhaes, A. L.; Cordeiro, M. N. D. S. Tetrahedron 2005, 61, 10951-10957; (d) Grieco, P. A.; Parker, D. T.; Fobare, W. F.; Ruckle, R. J. Am. Chem. Soc. 1987, 109, 5859-5861; (e) Grieco, P. A.; Larsen, S. D.; Fobare, W. F. Tetrahedron Lett. 1986, 27, 1975-1978; (f) Alonso, D. A.; Bertilsson, S. K.; Johnsson, S. Y.; Nordin, S. J. M.; Södergren, M. J.; Andersson, P. G. J. Org. Chem. 1999, 64, 2276-2280; (g) Bailey, P. D.; Wilson, R. D.; Brown, G. R. Tetrahedron Lett. 1989, 30, 6781-6784; (h) Waldman, H. Angew. Chem., Int. Ed. Engl. 1988, 27, 274-275; (i) Abraham, H.; Stella, L. Tetrahedron 1992, 48, 9707-9718; (j) Stella, L.; Abraham, H. Tetrahedron Lett. 1990, 31, 2603-2606.
- 14. Quadrelli, P.; Piccanello, A.; Vazquez Martinez, N.; Bovio, B.; Mella, M.; Caramella, P. Tetrahedron 2006, 62, 7370-7379.
- 15. (a) Daluge, S.; Vince, R. J. Org. Chem. 1978, 43, 2311-2320; (b) Katagiri, N.; Yamatoya, Y.; Ishikura, M. Tetrahedron Lett. 1999, 40, 9069-9072; (c) Evans, C. T.; Roberts, S. M.; Shoben, K. A.; Sutherland, A. G. J. Chem. Soc., Perkin Trans. 1 1992, 589-592.
- 16. (a) Katagiri, N.; Sato, H.; Kaneco, C.; Yusa, K.; Tsumo, T. J. Med. Chem. 1992, 35, 1882–1886; (b) Coe, D. M.; Myers, P. L.; Pony, D. M.; Roberts, S. M.; Stores, R. J. Chem. Soc. Chem. Commun. 1990, 151-153; (c) Evans, C. T.; Roberts, S. M.; Shoben, K. A.; Sutherland, A. G. J. Chem. Soc., Perkin Trans. 1 1992, 589-592.
- 17. Salvatori, D.; Volpini, R.; Vicenzetti, S.; Vita, A.; Costanzi, S.; Lambertucci, C.; Cristalli, G.; Vittori, S. Bioorg. Med. Chem. 2002, 10, 2973-2980.
- 18. (a) Corsico Coda, A.; Tacconi, G. Gazz. Chim. Ital. 1984, 114, 131-132; See also: (b) Grundmann, C.; Grünanger, P. The Nitrile Oxide: Versatile Tools of Theoretical and Preparative Chemistry; Springer: Heidelberg, 1971; (c) Torsell, K. J. B. Nitrile Oxides, Nitrones and Nitronates in Organic Synthesis; VCH: Weinheim, 1988.